[Product Name] Avian Influenza Panel III Nucle (AIV-H5, AIV-H7, AIV-H9) ic Acid Test Kit (Lyophilized)

[Package Specifications] 16 T/box

[Intended Use] This kit uses floorescence PCR methods to detect Newcastle disease Virus (NDV-U) in avian samples. This product requires operation with a real time quantitative PCR instrument and can achieve rapid POCT detection.

[Testing Principle] The test kit uses nucleic acid extraction reagents to extract the nucleic acid (DNA/RNA) from the sample. Under the acidon of a high-efficiency reverse transcriptase, cDNA complementary to the RNA template is synthesized in a one-step reaction using RNA as the template. Under the action of Taq enzyme, the copy number of the specific target fragment is amplified through cycles of high-temperature denaturation, annealing at a moderate temperature, and extension using DNA as the template. The fluorescence-labeled specific probe hybridizes with the amplified target fragment, and the 5'--3 exonuclease activity of Taq polymerase separates the reporting group an quencher group of the fluorescence probe, emitting a specific fluorescence signal. The specific fluorescence is detected using a fluorescence PCR instrument, and the result is determined based on the C1 value of the sample and the formation of the amplification curve.

[Contents]

Item	Quantity	Storage	
PCR master mix	16 pcs	-20°C (Away from light)	
Instructions for use	1 pcs	- Room Temperature	
Sample buffer	16 pcs		
Swab	16 pcs		
Biohazard bag	16 pcs		

[Storage conditions and shelf life] 1. Shelf life: 24 months. 2. Production date and expiration date e on the package.

[Compatible Instruments] This test kit is compatible with FLASHTEST real-time q fluorescence PCR instrument. antitati

[Sample]

cheal swab, fresh fecal

Casangle Handling]

Cloacal swab: insert the swab into the cloaca about 1.5~2cm, rotate at stain with feces.
Laryngotracheal swab: insert the swab from the mouth to the back of the pharynx directly to the larynx and trachea, wipe it gently and rotate it slowly, and stain it with tracheal secretions.
Fresh fecal: Use a swab to collect an appropriate amount.
With the swab in the sample buffer, shake it thoroughly to fully dissolve the pathogen on the swab head into the buffer.
Add 020 µL of mixed buffer to the nucleic acid extraction cartridge for extraction.

[Specimen storage] Samples used for nucleic acid extraction and detection should be tested as soon as possible. Samples to be tested within 24 hours can be stored at 4°C. Samples that can not be tested within 24 hours should be stored at -20°C for up to 10 days. Avoid repeated freezing and thawing of samples.

ction, to each PCR tub

[Instructions for Use] 1. Add Elution 1.1 Add 20µL of elution from magnetic bead extraction, to each PCR ti Close the lid tightly. 1.2 Shake all the liquid to the bottom of the PCR tube. Use the vortex mixer to mix the PCR tube horoughly, for 5 seconds. After mixing, ma sure all liquid is at the bottom of the PCR tube, by shaking the tube ag (optional: use a small centrifuge for 3 seconds to shift all liquids to the bottom.)

2. PCR Amplification 2.1 Set the parameters as follow

Step	Temperature	Time	Cycle
1	55°C	3min	1
2	94°C	30s	1
3	94°C 58°C	5s 20s	×40

2.2 The reaction volume is 20ul . Eluoresce nce channele

Channel	FAM	VIC	ROX	Cy5
Target	AIV-H7	Exogenous reference	AIV-H9	AIV-H5

3. Result Interpretation 3.1 Reference Range:

Parameter	Reference Range	Result Interpretation
Internal Control	Ct ≤ 37 and there is a clear exponential amplification curve	Valid
	Ct > 37 or No Ct	Invalid
Pathogen	Ct ≤ 37 and there is a clear exponential amplification curve	Positive
	Ct > 37 or No Ct	Negative

3.2 Test Result Interpretation

Pathogen Result	Internal Control Result	Test Result Interpretation
Positive	Valid	Pathogen Positive
Negative	Valid	Pathogen Negative
Any Result	Invalid	Test invalid, please retest

[Test Limitations] 1. The lest results of this kt should be comprehensively analyzed in conjunction with other relevant physical examination results and should not be used as the sole basis for diagnosis. 2. Improper sample collection, transportation, storage, handling, and inadequate laboratory conditions may lead to inaccurate results. 3. Other unconfirmed interferences or PCR inhibitors may lead to false negative results. 4. Sequence variations caused by mutations or other factors in the target gene of the virus being tested may lead to false negative results.

gene or the virus being tested may lead to false negative results. [Product Performanco] 1. Positive and negative control consistency: The positive and negative controls included in this test kit have been tested with the company's working reference materials, and the positive and negative compliance rates are both 100%. 2. Sensitivity: Imil of detection is 500 copies/mL. 3. Specificity: This assay does not cross-read with non-target pathoge samples. 4. Precision: The coefficient of variation (CV, %) of the Ct values for 10 consecutive tests of one strong positive sample and one weak positive sample is \leq %.

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