FIASHTEST

[Product Name] Avian Influenza H7 (AIV-H7)N ucleic Acid Test Kit (Lyophilize

(Package Speci

[Intended Use]
This kit uses fluorescence PCR methods to detect Avian Influenza H7
(AIV-H7) in avian samples. This product requires operation with a real tim
quantitative PCR instrument and can achieve rapid POCT detection.

quantitative PCR instrument and can achieve rapid POC1 detection.

[Testing Principle]
The test kit uses nucleic acid extraction reagents to extract the nucleic acid (DNA/RNA) from the sample.
Under the action of a high-efficiency reverse transcriptase, cDNA complementary to the RNA template is synthesized in a one-step reaction using RNA as the template. Under the action of Taq enzyme, the copy number of the specific larget fragment is amplified through cycles of high-temperature denaturation, annealing at a moderate temperature, and extension using DNA as the template. The fluorescence-labeled specific probe hybridizes with the amplified target fragment, and the 5'—3' exonuclease activity of Taq polymerase separates the reporting group and quencher group of the fluorescence probe, emitting a specific fluorescence signal. The specific fluorescence signal is detected using a fluorescence cosignal. The specific fluorescence PCR instrument, and the result is determined based on the Ct value of the sample and the formation of the amplification curve.

[Contents]			
Item	Quantity	Storage	
PCR master mix	16 pcs	-20°C (Away from light)	
Instructions for use	1 pcs	- Room Temperature	
Sample buffer	16 pcs		
Swab	16 pcs		
Biohazard bag	16 pcs		

[Storage conditions and shelf life 1. Shelf life: 24 months. 2. Production date and expiration of

[Compatible Instruments]
This test kit is compatible with FLASHTEST real-time quantitati fluorescence PCR instrument.

[Sample] Cloacal swab, laryngo icheal swab, fresh fecal

- [Sample Handling]
 1. Cloacal swab: insert the stain with feces.
- stain with feces.

 2. Laryngotracheal swab: insert the swab from the mouth to the back of the pharynx directly to the larynx and trachea, wipe it gently and rotate it slowly, and stain it with tracheal secretions.

 3. Fresh fecal: Use a swab to collect an appropriate amount.

 4.With the swab in the sample buffer, shake it thoroughly to fully dissolve the pathogen on the swab head into the buffer.

 5.Add 200 µL of mixed buffer to the nucleic acid extraction cartridge for

[Specimen storage]
Samples used for nucleic acid extraction and detection should be tested as soon as possible.
Samples to be tested within 24 hours can be stored at 4°C.
Samples that can not be tested within 24 hours should be stored at -20°C for up to 10 days.
Avoid repeated freezing and thawing of samples.

[Instructions for Use]

1. Add Elution

1. Add 2DuL of elution from magnetic bead extraction, to each PCR tube Close the lid tightly.

1.2 Shake all the liquid to the bottom of the PCR tube. Use the vortex mixer to mix the PCR tube thoroughly, for 5 seconds. After mixing, make sure all liquid is at the bottom of the PCR tube, by shaking the tube again. (optional: use a small centrifuge for 3 seconds to shift all liquids to the

2. PCR Amplification 2.1 Set the parameters as t

Step	Temperature	Time	Cycle
1	55°C	3min	1
2	94°C	30s	1
3	94°C 58°C	5s 20s	×40

2.2 The reaction volume is 20µL. Fluorescence channels:				
Channel	FAM	VIC	ROX	Cy5
Target	AIV-H7	Exogenous reference		

3. Result Interpretation

3.1 Reference Range:			
Parameter	Reference Range	Result Interpretation	
Internal Control	Ct ≤ 37 and there is a clear exponential amplification curve	Valid	
	Ct > 37 or No Ct	Invalid	
Pathogen	Ct ≤ 37 and there is a clear exponential amplification curve	Positive	
	Ct > 37 or No Ct	Negative	

3.2 Test Result Interpretation			
Pathogen Result	Internal Control Result	Test Result Interpretation	
Positive	Valid	Pathogen Positive	
Negative	Valid	Pathogen Negative	
Any Result	Invalid	Test invalid, please retest	

- [Test Limitations]

 1. The test results of this kit should be comprehensively analyzed in conjunction with other relevant physical examination results and should not be used as the sole basis for diagnosis.

 2. Improper sample collection, transportation, storage, handling, and inadequate laboratory conditions may lead to inaccurate results.

 3. Other unconfirmed interferences or PCR inhibitors may lead to false negative results.
- negative results.

 4. Sequence variations caused by mutations or other factors in the target gene of the virus being tested may lead to false negative results.

- [Product Performance]

 1. Positive and negative control consistency: The positive and negative controls included in this test kit have been tested with the company's working reference materials, and the positive and negative compliance rates are both 100%.

 2. Sensitivity: limit of detection is 500 copies/mL.

 3. Specificity: This assay does not cross-react with non-target pathogen samples.

 4. Precision: The coefficient of variation (CV, %) of the Ct values for 10 consecutive tests of one strong positive sample and one weak positive sample is ≤5%.

- sample is ≤5%.

 [Notes]

 1. Before using a PCR kit, check the lyophilized PCR mix at the bottom of the tube is in good condition (while and clumped). Liquiffed lyophilized PCR mix an not be used. After opening, it should be used as soon as possible or stored away from light.

 2. This product is only for in vitro testing (for animals). All operations mus strictly follow the instructions.

 3. Overloading samples may result in false negatives. Retest is recommended.

 4. Avoid bubbles in PCR tubes. Keep the tube cap firmly closed.

 5. Use disposable tips, gloves, and laboratory coats.

 6. After tests, disinfect the workbench with 10% hypocholrous acid, 75% ethanol, or UV light.

 7. All items in the kit should be treated as biowaste and handled in accordance with local laboratory regulations.