FIASHTEST

[Product Name] Canine Mycoplasma Nuc ic Acid Test Kit (Lyophil

[Package Specifications] 4 T/box

[Intended Use]
This kit uses fluorescence PCR methods to detect Canine Mycopeye, nose, and throat swab samples.
This product requires operation with a real time quantitative PCR instrument and can achieve rapid POCT detection.

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[Testing Principle]
The test kit uses nucleic acid extraction reagents to extract the nucleic acid (DNA/RNA) from the sample.
Under the action of a high-efficiency reverse transcriptase, cDNA complementary to the RNA template is synthesized in a one-step reauting RNA as the template.
Under the action of Tag enzyme, the copy number of the specific target fragment is amplified through cycles of high-temperature denaturation, annealing at a moderate temperature, and extension using DNA as the template.
The fluorescence-labeled specific probe hybridizes with the amplified target fragment, and the 5–3" exonuclease activity of Tag polymerase separates the reporting group and quencher group of the fluorescence probe, emitting a specific fluorescence signal. The specific fluorescence signal is detected using a fluorescence PCR instrument, and the result is determined based on the Ct value of the sample and the formation of the amplification curve.

| [Contents] | | | |
|----------------------|----------|----------------------------|--|
| Item | Quantity | Storage | |
| PCR master mix | 4 pcs | -20°C (Away from light) | |
| Instructions for use | 1 pcs | - Room Temperature | |
| Sample buffer | 4 pcs | | |
| Swab | 4 pcs | | |
| Biohazard bag | 4 pcs | | |

[Storage conditions and shelf life]
1. Shelf life: 24 months.
2. Production date and expiration date

ompatible Instruments] is test kit is compatible with FLASHTEST rea prescence PCR instrument.

[Sample] Eye, nose, and throat swab

- [Sample Handling]

 1. Eye, nose, and throat swab: Use a swab to moderately wipe the oral, nasal secretions, or conjunctival secretions.

 2. With the swab in the sample buffer, shake it thoroughly to fully dissolv the pathogen on the swab head into the buffer.

 3. add 200 µL of mixed buffer to the nucleic acid extraction cartridge for extraction.

[Specimen storage]
Samples used for nucleic acid extraction and detection should be tested as soon as possible.
Samples to be tested within 24 hours can be stored at 4°C.
Samples that can not be tested within 24 hours should be stored at -20°C for up to 10 days.
Avoid repeated freezing and thawing of samples.

[Instructions for Uso]

1. Add Elution

1. Add 20µL of elution from magnetic bead extraction, to each PCR tube. Close the lid tightly.

1.2 Shake all the liquid to the bottom of the PCR tube. Use the vortex mixe to mix the PCR tube thoroughly, for 5 seconds. After mixing, make sure all liquid is at the bottom of the PCR tube, by shaking the tube again. (optional: use a small centrifuge for 3 seconds to shift all liquids to the bottom.)

2. PCR Amplification 2.1 Set the parameters as follow

| Step | Temperature | Time | Cycle |
|------|--------------|-----------|-------|
| 1 | 55°C | 3min | 1 |
| 2 | 94°C | 30s | 1 |
| 3 | 94°C 58°C | 5s 20s | ×40 |

| Channel | FAM | VIC | CY5 | ROX |
|---------|------------|--------------------|-----|-----|
| Target | Mycoplasma | Internal reference | | |

| 1. Reference Range: | | | |
|---------------------|--|--------------------------|--|
| Parameter | Reference Range | Result Interpretation | |
| Internal Control | Ct ≤ 37 and there is a clear exponential amplification curve | Valid | |
| | Ct > 37 or No Ct | Invalid | |
| Pathogen | Ct ≤ 37 and there is a clear exponential amplification curve | Positive | |
| | Ct > 37 or No Ct | Negative | |

| 3.2 Test Result Interpretation | | | |
|--------------------------------|----------------------------|-------------------------------|--|
| Pathogen Result | Internal Control Result | Test Result Interpretation | |
| Positive | Valid | Pathogen Positive | |
| Negative | Valid | Pathogen Negative | |
| Any Result | Invalid | Test invalid, please retest | |

[Test Limitations]

1. The lest results of this kit should be comprehensively analyzed in conjunction with other relevant physical examination results and should not be used as the sole basis for diagnosis.

2. Improper sample collection, transportation, storage, handling, and inadequate laboratory conditions may lead to inaccurate results.

3. Other unconfirmed interferences or PCR inhibitors may lead to false negative results.

4. Sequence variations caused by mutations or other factors in the targegene of the virus being tested may lead to false negative results.

[Product Performance]

1. Positive and negative control consistency. The positive and negative control is included in this test kit have been tested with the company's working reference materials, and the positive and negative compliance rates are both 100%.

2. Sensitivity: limit of detection is 500 copies/mL.

3. Specificity. This assay does not cross-react with non-target pathoge samples.

4. Precision: The coefficient of variation (CV, %) of the Ct values for 10 consecutive tests of one strong positive sample and one weak positive sample is ≤5%.

- Notes]

 1. Before using a PCR kit, check the lyophilized PCR mix at the bottom of the tube is in good condition (white and clumped). Liquified lyophilized PCR mix can not be used. After opening, it should be used as soon as PCR mix can not be used. After opening, it should be used as soon as soon as a soo