

[Product Name] Feline Anemia IV Nucleic Acid Test Kit (Lyophilized) (FeLV, FIV, M. hemophilic, B. henselae)

[Package Spe ons]

[Intended Use] This kit uses fluorescence PCR methods to detect Feline Leukemia virus (FeLV), Feline Immunodeficiency virus (FIV), Mycoplasma haemofelis (M. hemophilic), and Bartonelia henselae (B. henselae) in biood sample. This product requires operation with a real time quantitative PCR instrument and can achieve rapid POCT detection.

Instrument and can achiever rapid POCT detection. [resting Principle] The test kit uses nucleic acid extraction reagents to extract the nucleic acid (DNA/RNA) from the sample. Under the acition of a high-efficiency reverse transcriptase, cDNA complementary to the RNA template is synthesized in a one-step react using RNA as the template. Under the acition of Tag enzyme, the copy number of the specific target fragment is amplified through cycles of high-temperature denaturation, annealing at a moderate temperature, and extension using DNA as the template. The fluorescence-labeled specific probe hybridizes with the amplified target fragment, and the G⁻³ exonuclease activity of Tag polymerase separates the reporting group and quencher group of the fluorescence PCR The specific fluorescence signal is detected using a fluorescence PCR instrument, and the formation of the amplification curve.

[Conte 4.1

Item	Quantity	Storage
PCR master mix	4 pcs	-20°C (Away from light)
Instructions for use	1 pcs	
Sample buffer	4 pcs	Room Temperature
Biohazard bag	4 pcs	

[Storage conditions and shelf life] 1. Shelf life: 24 months. 2. Production date and expiration date are on the package.

[Compatible Instruments] This test kit is compatible with FLASHTEST r fluorescence PCR instrument.

[Sample] EDTA anticoagulated blood

[Sample Handling] EDTA anticoagulated blood: Collect blood using a blood collection tube containing EDTA anticoagulant. 1. Add 100 µL of blood to the sample buffer with a disposable dropper. 2. Thoroughly mix the sample buffer with a repetitive pipetting action, using the disposable dropper. 3. Add 200 µL of mixed buffer to the nucleic acid extraction cartridge for

[Specimen storage] Samples used for nucleic acid extraction and detection should be tested as soon as possible. Samples to be tested within 24 hours can be stored at 4°C. Samples that can not be tested within 24 hours should be stored at -20°C for up to 10 days. Avoid repeated freezing and thawing of samples.

[Instructions for Use] 1. Add Elution 1. Add ZoluL of elution from magnetic bead extraction, to each PCR tube Close the lid tightly. 1.2 Shake all the liquid to the bottom of the PCR tube. Use the vortex mixer to mix the PCR tube thoroughly, for 5 seconds. After mixing, make sure all liquid is at the bottom of the PCR tube, by shaking the tube again. (optional: use a small centrifuge for 3 seconds to shift all liquids to the bottom.)

2. PCR Amplification 2.1 Set the parameters as follo

Step	Temperature	Time	Cycle
1	55°C	3min	1
2	94°C	30s	1
3	94°C 58°C	5s 20s	×40

2.2 The reaction volume is 20µL. Fluorescence channels:

onumer	1.000	10	010	NOA
Target (Tube 1)	FeLV	Internal reference	FIV	
Target (Tube 2)	M. hemophilic			B. henselae

3. Result Interpretation 3.1 Reference Range:

Parameter	Reference Range	Result Interpretation
Internal Control	Ct ≤ 37 and there is a clear exponential amplification curve	Valid
	Ct > 37 or No Ct	Invalid
Pathogen	Ct ≤ 37 and there is a clear exponential amplification curve	Positive
	Ct > 37 or No Ct	Negative

3.2 Te t Result Interpretation

Pathogen Result	Internal Control Result	Test Result Interpretation
Positive	Valid	Pathogen Positive
Negative	Valid	Pathogen Negative
Any Result	Invalid	Test invalid, please retest

[Test Limitations]
1. The test results of this kit should be comprehensively analyzed in conjunction with other relevant physical examination results and should not be used as the sole basis for diagnossi.
2. Improper sample collection, transportation, storage, handling, and inadequate laboratory conditions may lead to inaccurate results.
3. Other unconfirmed interferences or PCR inhibitors may lead to false negative results.
4. Sequence variations caused by mutations or other factors in the targe gene of the virus being tested may lead to false negative results.

gene on the whos being tested may read to take negative results. [Product Performance] 1. Positive and negative control consistency: The positive and negative controls included in this test kit have been tested with the company's working reference materials, and the positive and negative compliance rates are both 100%. 3. Specificity: This assay does not cross-read with non-target pathoge samples. 4. Precision: The coefficient of variation (CV, %) of the Ct values for 10 consecutive tests of one strong positive sample and one weak positive sample is ≤5%.

[Notes] 1. Before

[Notes] 1. Before using a PCR kit, check the lyophilized PCR mix at the bottom of the tube is in good condition (while and clumped). Liquified lyophilized PCR mix can not be used. After opening, it should be used as soon as possible or stored away from light. 2. This product is only for in vitro testing (for animals). All operations mus strictly follow the instructions. 3. Overloading samples may result in false negatives. Retest is recommended. 4. Avoid bubbles in PCR tubes. Keep the tube cap firmly closed. 5 Lise disnosable tips, cloves, and laboratory coats.

recommended. 4. Avoid hubbes in PCR tubes. Keep the tube cap firmly closed. 5. Use disposable tips, gloves, and laboratory coats. 6. After tests, disinfect the workbench with 10% hypochlorous acid, 75% ethanol, or UV light. 7. All items in the kit should be treated as biowaste and handled in accordance with local laboratory regulations.