### **FIASHTEST**

[Product Name]
Feline Respiratory Panel V Nucleic Acid Test Kit(Lyophiliz (FHV, FCV, Mycoplasma, Chlamydia, B. bronchiseptica)

## [Package Spe

[Intended Use]
This kit uses fluorescence PCR methods to defect FHV, FCV, Mycoplasma
Chlamydia and B. bronchiseptica in eye, nose, and throat swab samples.
This product requires operation with a real time quantitative PCR instrumer
and can achieve rapid PCCT detection.

and can achieve rapid POCT detection.

[Testing Principle]
The test kit uses nucleic acid extraction reagents to extract the nucleic acid (DNA/RNA) from the sample.
Under the action of a high-efficiency reverse transcriptase, cDNA complementary to the RNA template is synthesized in a one-step reaction using RNA as the template.
Under the action of Tag enzyme, the copy number of the specific target fragment is amplified through cycles of high-temperature denaturation, annealing at a moderate temperature, and extension using DNA as the template.
The fluorescence-labeled specific probe hybridizes with the amplified target fragment, and the 5"--3" exonuclease activity of Tag polymerase separates the reporting group and quencher group of the fluorescence probe, emitting a specific fluorescence signal is detected using a fluorescence PCR instrument, and the result is determined based on the Ct value of the sampli and the formation of the amplification curve.

[Contents]			
Item	Quantity	Storage	
PCR master mix	4 pcs	-20°C (Away from light)	
Instructions for use	1 pcs		
Sample buffer	4 pcs	Room Temperature	
Swab	4 pcs	Room remperature	
Biohazard bag	4 pcs		

[Storage conditions and shelf life]
1. Shelf life: 24 months.
2. Production date and expiration date

# [Compatible Instruments] This test kit is compatible with FLASHTEST refluorescence PCR instrument.

## [Sample] Eye, nose, and thro

[Sample Handling]

1. Eye, nose, and throat swab: Use a swab to moderately wipe the oral, nasal secretions, or conjunctival secretions.

2. With the swab in the sample buffer, shake it thoroughly to fully dissolvant pathogen on the swab head into the buffer.

3. add 200 µL of mixed buffer to the nucleic acid extraction cartridge for extraction.

[Specimen storage]
Samples used for nucleic acid extraction and detection should be tested as soon as possible.
Samples to be tested within 24 hours can be stored at 4°C.
Samples that can not be tested within 24 hours should be stored at -20°C for the total control of the stored at -20°C for the total control of the stored at -20°C for the total control of the stored at -20°C for the total control of the stored at -20°C for the stored

[Instructions for Uso]

1. Add Elution

1.1 Add 20 Ju. of elution from magnetic bead extraction, to each PCR tube. Close the lidt lightly.

1.2 Shake all the liquid to the bottom of the PCR tube. Use the vortex mixe to mix the PCR tube throughly, for 5 seconds. After mixing, make sure all liquid is at the bottom of the PCR tube, by shaking the tube again. (optional: use a small centrifuge for 3 seconds to shift all liquids to the bottom.)

Step	Temperature	Time	Cycle
1	55°C	3min	1
2	94°C	30s	1
3	94°C 58°C	5s 20s	×40

Channel	FAM	VIC	CY5	ROX
Target (Tube 1)	FCV	Internal reference	FHV	
Target (Tube 2)	Mycoplasma		Chlamydia	B. bronchiseptica

3.1 Reference Range:			
Parameter	Reference Range	Result Interpretation	
Internal Control	Ct ≤ 37 and there is a clear exponential amplification curve	Valid	
	Ct > 37 or No Ct	Invalid	
Pathogen	Ct ≤ 37 and there is a clear exponential amplification curve	Positive	
	Ct > 37 or No Ct	Negative	

0.2 Test Nesult Interpretation			
Pathogen Result	Internal Control Result	Test Result Interpretation	
Positive	Valid	Pathogen Positive	
Negative	Valid	Pathogen Negative	
Any Result	Invalid	Test invalid, please retest	

- [Test Limitations]

  1. The lest results of this kit should be comprehensively analyzed in conjunction with other relevant physical examination results and should not be used as the sole basis for diagnosis.

  2. Improper sample collection, transportation, storage, handling, and inadequate laboratory conditions may lead to inaccurate results.

  3. Other unconfirmed interferences or PCR inhibitors may lead to false negative results.

  4. Sequence variations caused by mutations or other factors in the targe gene of the virus being tested may lead to false negative results.

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  Precision: The coefficient of variation (CV, %) of the Ct values for 10 onsecutive tests of one strong positive sample and one weak positive ample is ≤5%.

- sample is \$50%.

  [Notes]

  1. Before using a PCR kit, check the lyophilized PCR mix at the bottom of the tube is in good condition (white and clumped). Liquiffed lyophilized PCR mix an not be used. After opening, it should be used as \$500 as \$500